

Table 4.1
Summary of findings.

Type of samples	Outcome	Number of included studies with low or moderate risk of bias	Comments
NGS for targeted analysis			
NIPT	Monosomy X	9	Few events, inconsistency in data, differences in study design, population and sequencing method applied
NIPT	XXX	4	Few events, inconsistency in data, differences in study design, population and sequencing method applied
NIPT	XXY	5	Few events, inconsistency in data, differences in study design, population and sequencing method applied
NIPT	XYY	4	Few events, inconsistency in data, differences in study design, population and sequencing method applied
NIPT	Trisomies other than T13, T18, T21	3	8 cases of other autosomal trisomies
NIPT	Microdeletions associated with known microdeletion syndromes	1 (2 studies with high risk of bias)	Only cases identified with NGS underwent invasive sampling for confirmation
NIPT	Microduplications associated with known microduplication syndromes	0 (0 studies with high risk of bias)	No studies identified
NGS for whole genome sequencing/whole exome sequencing			
NIPT		2 (0 studies with high risk of bias)	Few events with different genetic abbreviations. Different methods applied in the two studies
Invasive		0 (3 studies with high risk of bias)	Highly selected populations. Only studies with high risk of bias identified.

NGS = Next-generation sequencing; **NIPT** = Non-invasive prenatal testing

Table 4.2 Included studies investigating NIPT with NGS for identification of sex chromosomal aneuploidies (SCA).

Author, year Reference Country	Study design	Sequencing method	Time of study	Population Risk for aneuploidy Mean maternal age (years) Mean gestational age (weeks)	Outcome	Number of included samples Number of samples lost to follow-up
Jiang, 2012 [30] China	Prospective, multicenter (biobanked samples)	MPSS	2009–2010	High risk 20–45 10–34	SCA	903 1
Liang, 2013 [31] China	Prospective, multicenter (biobanked samples)	Not specified	2009–2011	High risk 31 21	SCA and T9	435 23
Porreco, 2014 [29] USA	Prospective, multicenter, (biobanked samples)	MPSS	2009–2011	High risk 35 16	SCA	4 170 969 (892) (different number of non- reportable result for different outcomes)
Bianchi, 2012 [22] USA	Prospective, multicenter, nested case control (biobanked samples)	MPSS	2010–2011	High risk 35 15	Monosomy X	532 (selected from 2 882 samples) 2
Song, 2013 [25] China	Prospective cohort (not biobanked samples)	Not specified	2011	High and low risk 29 (all less than 35) 17	SCA (not all samples underwent invasive reference test)	1 916 184
Yao, 2014 [27] China	Prospective cohort, (not biobanked samples)	Not specified	2011–2012	High and low risk 30 19	SCA (not all samples underwent invasive reference test)	5 950 484
Lau, 2014 [26] China	Prospective cohort, (not biobanked samples)	Not specified	2011–2013	High and low risk 36 14,5 (median)	SCA + other trisomies, deletions and duplications (not all samples underwent invasive reference test)	1 982 1

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Table 4.2 continued

Author, year Reference Country	Study design	Sequencing method	Time of study	Population Risk for aneuploidy Mean maternal age (years) Mean gestational age (weeks)	Outcome	Number of included samples Number of samples lost to follow-up
Song, 2015 [24] China	Prospective cohort (not biobanked samples)	Not specified	2012–2013	High risk 37 (all over 35) 10 (only first trimester)	SCA	213 35
Shaw, 2014 [34] China	Prospective, multicenter cohort (not biobanked samples)	MPSS	2012	High and low risk 35 17	SCA and T16	201 1
Comas, 2015 [28] Spain	Prospective cohort (not biobanked samples)	SNP/ t-MPS	2013	High and low risk 37 15	SCA (only high risk pregnancies at NIPT underwent the invasive reference test)	333 (217 analysed for SCA) 22
Mazloom, 2013 [32] USA	Prospective cohort (biobanked samples)	MPSS	ns	High risk 36 (median) 17 (median)	SCA	411 21
Nicolaides, 2013 [33] United Kingdom	Prospective cohort (biobanked samples)	SNP	ns	High risk 36 (median) 13 (median)	SCA	242 13
Pergament, 2014 [17] USA	Prospective, multicenter cohort, (not biobanked samples)	SNP	ns	High and low risk 30 14	SCA	1 064 98

ns = Not stated; MPSS = Massive parallel shotgun sequencing; SCA = Sex chromosome aneuploidy; SNP = Single nucleotide polymorphism; T = Trisomi; t-MPS = Targeted massive parallel sequencing

Figur 4.1-4.4 Sensitivitet och specificitet för NIPT med NGS för att identifiera könskromosomavvikelser.

Figure 4.1 Sensitivitet och specificitet för identifikation av monosomi X.

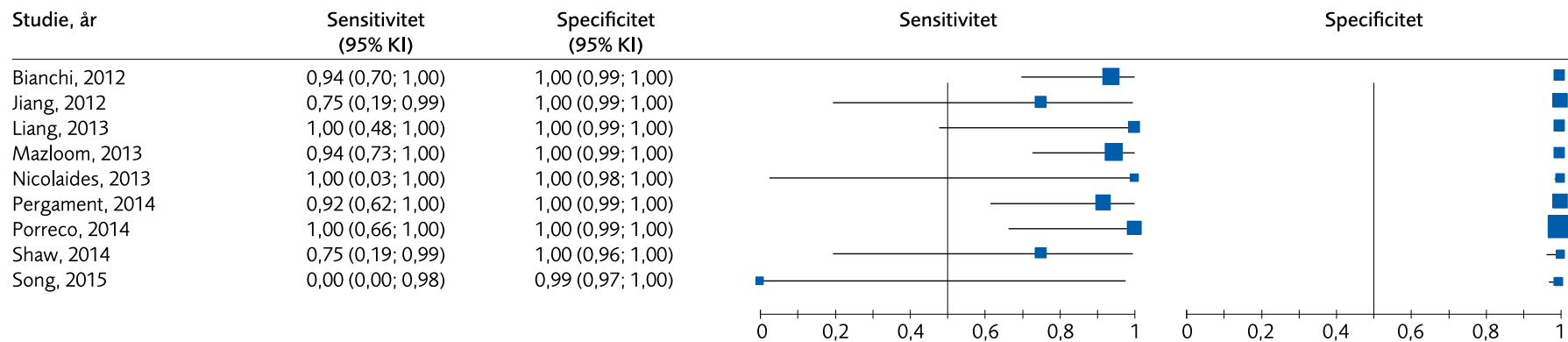


Figure 4.2 Sensitivitet och specificitet för identifikation av XXX.

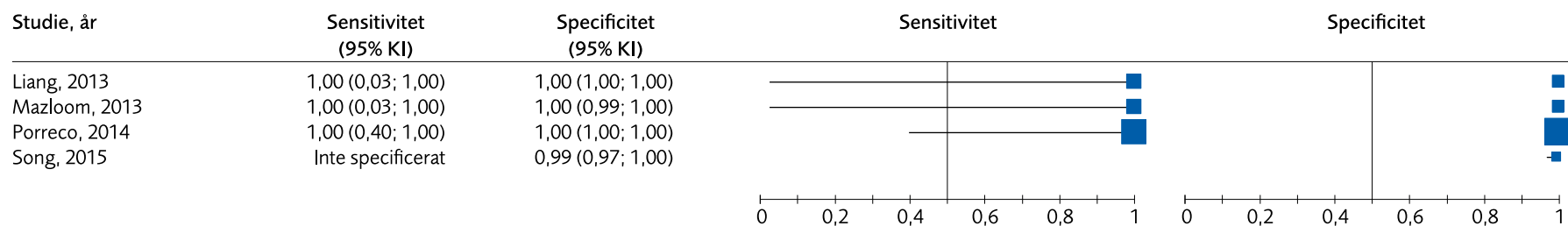


Figure 4.3 Sensitivitet och specificitet för identifikation av XXY.

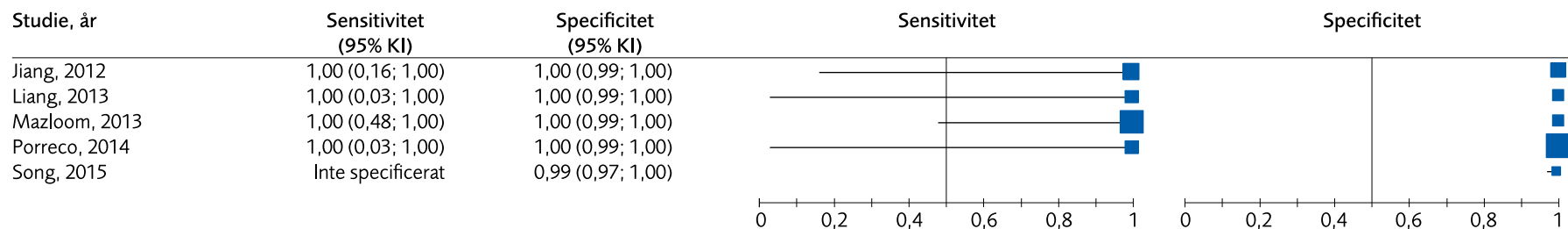


Figure 4.4 Sensitivitet och specificitet för identifikation av XYY.

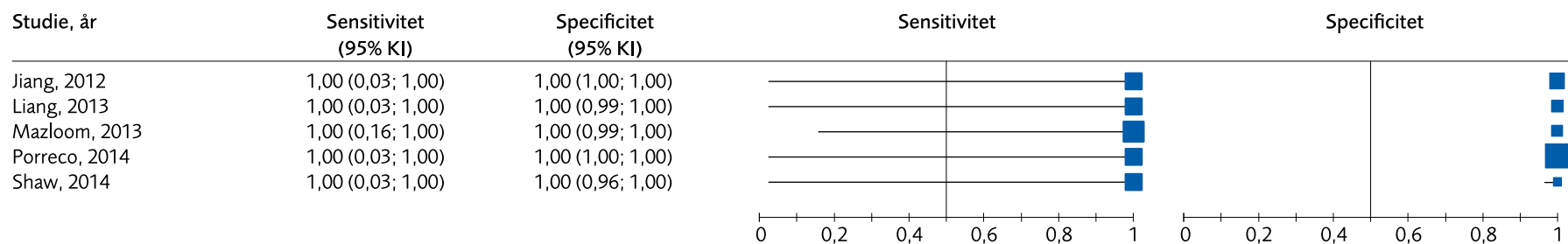


Table 4.3 Distribution of positive events in studies using NGS and non-invasive samples (NIPT).

Outcome	Number of included studies	Only true positive events: Number of studies Number of events (number of successful analyses)	Only false positive events: Number of studies Number of events (number of successful analyses)	Both true and false positive events: Number of studies Number of events (number of successful analyses)
Monosomy X	13	4 7 (332)	2 2 (179)	7 81 (6 379)
XXX	6	3 4 (804)	1 1 (178)	2 17 (3 280)
XXY	7	3 8 (1 705)	2 2 (179)	2 10 (3 208)
YY	6	5 6 (5 006)	0	1 1 (2*)

* Only cases with positive NIPT results underwent confirmation from invasive sampling.

Table 4.4 Number of analyses, true and false positive events, and the proportion of false positive events in the studies where both true and false positive event were identified.

Outcome	Number of Studies	Total number of successful analyses	Number of true positive events Median (range)	Number of false positive events Median (range)	Proportion of false positive events Median (range)
Monosomy X	7	6 379	9 (3–17)	1 (1–11)	16.7% (6.3–55%)
XXX	2	3 280	5.5 (4–7)	3 (3–3)	36.4% (30.0–42.9%)
XXY	2	3 208	2 (1–3)	3 (2–4)	61.9% (57.1–66.7%)
YY	1	2	1	1	50%

Table 4.5 Distribution of negative events in studies using NGS and non-invasive samples (NIPT).

Outcome	Number of included studies	Only true negative events: Number of studies (Total number of successful analyses)	Only false negative events	Both true and false negative events: Number of studies Number of false negative events (Number of successful analyses)
Monosomy X	9	3 (3 911)	0	6 5 (2 870)
XXX	4	4 (4 250)	0	0
XXY	5	5 (5 084)	0	0
XYY	5	5 (5 006)	0	0

Table 11.1 Included studies Next-generation sequencing from NIPT sample to detect sex chromosome aneuploidies and trisomies other than T13, T18 and T21.

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Bianchi 2012 [22] USA	<p>Study design Nested case control study within a prospective multicenter, observational cohort</p> <p>Blinded</p> <p>Time of study June 2010 to August 2011</p>	<p>Population High risk for fetal aneuploidy n=2 882 (cohort) n=534 (selected) Successful samples n=433</p> <p>Samples Biobanked samples</p> <p>Inclusion criteria Age 38 years or older Prior aneuploid pregnancy Anomaly detected by USS Positive maternal serum screen</p> <p>Exclusion criteria Ineligible samples, no karyotype recorded or multiple gestation</p> <p>Gestational age at sampling 10–23 weeks</p> <p>Maternal age Mean 35 years</p> <p>Drop-outs Did not pass quality control n=2 No fetal DNA detected n=16 Censored complex karyotype n=37 Unclassified =49 (when checked number should read 46)</p>	<p>Platform NIPT MPSS</p> <p>Reference test Karyotype</p>	<p>Monosomy X TP 15 FP 1 FN 1 TN 416</p> <p>Sensitivity: 93.8 (95% CI, 69.8; 99.8) Specificity: 99.8 (95% CI, 98.7; 99.9)</p>	<p>Moderate</p> <p>Commercial partner Verinata</p>

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First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Comas 2015 [28] Spain	<p>Study design Prospective cohort study</p> <p>Not blinded</p> <p>Time of study January to December 2013</p>	<p>Population Mainly low risk for fetal aneuploidy (84%) n=333 (total) n=311 (analysed)</p> <p>Samples</p> <p>Not biobanked</p> <p>Inclusion criteria Singleton pregnancies Pregnancies referred by anxiety or high risk in maternal serum screen combined with nuchal translucency</p> <p>Exclusion criteria Anomaly detected by USS Pregnancies at high risk of other genetic conditions</p> <p>Gestational age at sampling 9–23 weeks</p> <p>Maternal age Mean 37 years (21–46)</p> <p>Drop-outs No test result n=4 Repeated sampling and analysis n=6 Lost to follow up n=18 (pregnancies that were still in progress)</p>	<p>Platform NIPT SNP/T-MPS</p> <p>Reference test Karyotype or neonatal data</p> <p>Karyotyping only in patients with high risk in NIPT analysis</p>	<p>Monosomy X (1 identified with NIPT, 1 analysed with karyotype) TP 0 FP 1 Lost to follow-up: 0</p>	<p>Moderate</p> <p>Commercial partner Natera</p>

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Jiang 2012 [30] China	Study design Multicenter prospective cohort study Blinding unclear Time of study June 2009 to August 2010	Population High risk for fetal aneuploidy n=903 (total) n=903 (analysed) Samples Biobanked samples Inclusion criteria Pregnant women undergoing invasive test Exclusion criteria Not specified Gestational age at sampling 10–34 weeks Maternal age Range 20–45 years Drop-outs 1 due to failure in gender classification (calculated as ITT)	Platform NIPT MPSS Reference test Karyotype	Monosomy X TP 3 FP 1 FN 1 TN 898 Sensitivity: 75% Specificity: 99.9% XXY TP 2 FP 0 FN 0 TN 901 Sensitivity: 100% Specificity: 100% XYY TP 1 FP 0 FN 0 TN 902 Sensitivity: 100% Specificity: 100%	Moderate Commercial partner BGI

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Lau 2014 [26] Hong Kong	<p>Study design Prospective cohort single center study</p> <p>Blinded</p> <p>Time of study August 2011 to February 2013</p>	<p>Population n=1 982 (total) n=1 981 (analysed)</p> <p>Samples Not biobanked samples 1 929 singleton, 30 twin fetuses, 23 internal controls (anomaly detected by USS)</p> <p>Inclusion criteria Physician request of NIPT, pregnancy >12 weeks</p> <p>Exclusion criteria Major anomaly detected by USS</p> <p>Gestational age at sampling Median 14.5 weeks</p> <p>Maternal age Mean 36 years (20–46)</p> <p>Drop-outs Failed analysis n=1 Repeat blood sample n=23 Lost to follow-up 14.4%</p>	<p>Platform NIPT Not specified</p> <p>Reference test Karyotype, QF-PCR or CMA (only NIPT positive & on 23 internal controls)</p> <p>NIPT negative cases were contacted within 3 months after expected delivery for a clinical outcome follow up</p>	<p>Monosomy X (1 identified with NIPT, 1 analysed with karyotype) TP 1 FP 0 Lost to follow-up: 0</p> <p>XXX (4 identified with NIPT, 2 analysed with karyotype) TP 2 FP 0 Lost to follow-up: 2</p> <p>XXY/XXY (3 identified with NIPT, 2 analysed for karyotype) TP 2 FP 0 Lost to follow-up: 1</p> <p>Autosomal trisomy 4/6 detected trisomies with NIPT were analysed with karyotyping TP 0 FP 4</p> <p>Triple trisomy 1 detected with NIPT but confirmed confined to placental mosaicism</p> <p>Deletion/duplication 1 partial trisomy 18 1 partial monosomy 18 Confirmed for fetal or maternal origin</p>	<p>Moderate</p> <p>Commercial interests BGI</p>

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Liang 2013 [31] China	Study design Multicenter prospective cohort study Blinded Time of study March 2009 to June 2011	Population High risk for fetal aneuploidy n=435 (total) n=412 (analysed) Samples Biobanked samples Inclusion criteria Positive maternal serum screen n=217 Anomaly detected by USS n=67 Prior aneuploidy pregnancy n=4 AMA ≥ 35 n=84 More than one indication n=63 Exclusion criteria Not specified Gestational age at sampling Median 21 weeks (11–39 weeks) Maternal age Mean 31 years Drop-outs n=11 karyotypes missing n=12 failed sequencing	Platform NIPT Not specified Reference test Karyotype	Monosomy X (3 mosaic 0X treated as diploid: 1 detected and 2 mosaic missed in sequencing) TP 5 FP 1 FN 0 TN 406 Sensitivity: 100% Specificity 99.8% XXX TP 1 FP 0 FN 0 TN 411 Sensitivity and Specificity: 100% XXY TP 1 FP 0 FN 0 TN 411 Sensitivity and Specificity: 100% XYY TP 1 FP 0 FN 0 TN 411 Sensitivity and Specificity: 100% Trisomy 9 TP 1 FP 0 FN 0 TN 411 Sensitivity and Specificity: 100%	Moderate Commercial partner Berry Genomics

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Mazloom 2013 [32] USA	Study design Prospective cohort study Blinded Time of study Not reported	Population High risk for fetal aneuploidy Validation set only population n=411 Successful samples n=390 Samples Biobanked samples Inclusion criteria Invasive sampling for karyotyping due to high risk for fetal aneuploidy Exclusion criteria Multiple gestation, mosaicism for sex chromosomes; no documented karyotype reported Gestational age at sampling 8–29 weeks Maternal age Median 36 years (19 to 47) Drop-outs Non reportable results n=21 (5%)	Platform NIPT MPSS Reference test Karyotype/FISH or both	Monosomy X TP 17 FP 1 FN 1 TN 371 XXX TP 1 FP 0 FN 0 TN 389 XXY TP 5 FP 0 FN 0 TN 385 XYY TP 2 FP 0 FN 0 TN 388	Moderate Commercial partner Sequenom Inc

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First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Nicolaidis 2013 [33] United Kingdom	Study design Prospective cohort study Blinded Time of study Not reported	Population High risk for fetal aneuploidy n=242 (total) n=229 (analysed) Samples Biobanked samples Inclusion criteria AMA Prior aneuploid pregnancy Positive maternal serum screen Exclusion criteria Not specified Gestational age at sampling 11–14 weeks Maternal age Median 36 years (18–47) Drop-outs n=13 failed quality control	Platform NIPT SNP Reference test Karyotype	Monosomy X TP 2 FP 0 FN 0 TN 227 Triploidy TP 1 FP 0 FN 0 TN 228	Moderate Commercial partner None reported (personal communication)

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Pergament 2014 [17] USA	<p>Study design Prospective cohort multicenter study</p> <p>Blinded</p> <p>Time of study Not reported</p>	<p>Population n=1 064 (total) n=966 (analysed)</p> <p>High risk for fetal aneuploidy n=543 (total) n=492 (analysed, mosaic monosomy X cases included)</p> <p>Low risk for fetal aneuploidy n=521 (total) n=474 (analysed, mosaic monosomy X cases included)</p> <p>Samples Not biobanked samples</p> <p>Inclusion criteria >18 years Singleton pregnancy >7 weeks</p> <p>Exclusion criteria Confirmed sex chromosome aneuploidy, triploidy or fetal mosaicism</p> <p>Gestational age at sampling Median 14 weeks (7–40)</p> <p>Maternal age Median 30 years (18–47)</p> <p>Drop-outs Did not pass quality control n=85 Fetal mosaicism monosomy X n=2</p>	<p>Platform NIPT SNP</p> <p>Reference test Karyotype or FISH</p>	<p>Monosomy X TP 9 FP 1 FN 1 TN 953</p> <p>Sensitivity: 90 (95% CI, 55.5; 99.8) Specificity: 99.9 (95% CI, 99.4; 100)</p> <p>Monosomy X including mosaic karyotypes TP 11 FP 1 FN 1 TN 953</p> <p>Sensitivity: 91.7 (95% CI, 61.5; 99.8) Specificity: 99.9 (95% CI, 99.4; 100)</p>	<p>Moderate</p> <p>Commercial partner Natera</p>

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Porreco 2014 [29] USA	<p>Study design Prospective multicenter observational study</p> <p>Blinded</p> <p>Time of study September 2009 to April 2011</p>	<p>Population High risk for fetal aneuploidy n=4 170 (total), n= 3 430 (selected) Successful samples for outcome of X or XXX n=3 278 Successful samples for outcome of XXY or XYY n=3 201</p> <p>Samples Biobanked samples</p> <p>Inclusion criteria AMA, Personal or family history Positive maternal serum screen Anomaly detected by USS , Singleton pregnancy ≥18 years of age</p> <p>Exclusion criteria Inability to give written informed consent, multiple gestation, fetal demise of additional embryo during current pregnancy Insufficient sample volume Outside time window for laboratory processing</p> <p>Gestational age at sampling Mean 16 weeks (9–37)</p> <p>Maternal age Mean 35 years (18–50)</p> <p>Drop-outs Low quantity of fetal DNA or complex karyotypes: Outcome X or XXX n=152 (4.6%) Outcome XXY or XYY n=229 (7.1%)</p>	<p>Platform NIPT MPSS</p> <p>Reference test Karyotyping</p>	<p>Monosomy X TP 9 FP 11 FN 0 TN 3 258</p> <p>Sensitivity: 100 (95% CI, 66.4; 100) Specificity: 99.7 (95% CI, 99.4; 99.8)</p> <p>XXX TP 4 FP 3 FN 0 TN 3 271</p> <p>Sensitivity: 100 (95% CI, 39.8; 100) Specificity: 99.9 (95% CI, 99.7; 100)</p> <p>XXY TP 1 FP 2 FN 0 TN 3 198</p> <p>Sensitivity: 100 (95% CI, 2.50; 100) Specificity: 99.9 (95% CI, 99.8; 100)</p> <p>XYY TP 1 FP 0 FN 0 TN 3 200</p> <p>Sensitivity: 100 (95% CI, 2.5; 100) Specificity: 100 (95% CI, 99.9; 100)</p>	<p>Moderate</p> <p>Commercial partner Sequenom</p>

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Shaw 2014 [34] Taiwan	<p>Study design Prospective multi-center cohort study</p> <p>Blinded</p> <p>Time of study June to December 2012</p>	<p>Population n=201 (total) n=200 (analysed)</p> <p>High risk n=100 (total and analysed)</p> <p>Low risk n=101 (total) n=100 (analysed)</p> <p>Samples Not biobanked samples</p> <p>Gestational age at sampling High risk: Mean 17 weeks Low risk: Mean 16 weeks</p> <p>Inclusion criteria High risk: Risk of 1:30, NT >3.0 mm Low risk: Risk of 1:1 500</p> <p>Exclusion criteria Not specified</p> <p>Maternal age High risk: Mean 35 years Low risk: Mean 35 years</p> <p>Drop-outs Due to early gestational age n=1</p>	<p>Platform NIPT MPSS</p> <p>Reference test Karyotype and phenotype at birth</p> <p>In the high risk group all samples were analysed with karyotype</p>	<p>High risk group Monosomy X TP 3 FP 0 FN 1 TN 96</p> <p>XYY TP 1 FP 0 FN 0 TN 99</p> <p>Trisomi 16 TP 1 FP 0 FN 0 TN 99</p> <p>Low risk group 45X, 47XYY or Trisomi 16 TP 0 FP 0 FN 0 TN 100</p>	<p>Moderate</p> <p>Commercial partner Berry genomics</p>

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Song 2013 [25] China	<p>Study design Prospective cohort study</p> <p>Not Blinded</p> <p>Time of study April to December 2011</p>	<p>Population High risk for fetal aneuploidy n=1 916 (total) n=1 741 (analysed by DNA sequencing) n=202 (karyotype)</p> <p>Samples Not biobanked samples</p> <p>Inclusion criteria Singleton pregnancies Maternal age less than 35 years Positive maternal serum screen</p> <p>Exclusion criteria Not specified</p> <p>Gestational age at sampling Mean 17 weeks (11–21)</p> <p>Maternal age Mean 29 years</p> <p>Drop-outs Failed sequencing quality control n=73 Lost to follow-up, birth outcome n=111</p>	<p>Platform NIPT Not specified</p> <p>Reference test Karyotyping</p>	<p>Monosomy X (2 identified with NIPT, 2 analysed with karyotype) TP 2 FP 0 Lost to follow-up: 0</p> <p>XXY (1 identified with NIPT, 1 analysed with karyotype) TP 0 FP 1 Lost to follow-up: 0</p>	<p>Moderate</p> <p>Commercial partner None reported</p>

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Table 11.1 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Song 2015 [24] China	<p>Study design Prospective cohort study</p> <p>Blinded</p> <p>Time of study May 2012 to August 2013</p>	<p>Population High risk for fetal aneuploidy n=213 (total) n=178 (analysed by both methods)</p> <p>Samples Not biobanked samples</p> <p>Inclusion criteria AMA >35 years Singleton pregnancy</p> <p>Exclusion criteria Not specified</p> <p>Gestational age at sampling 8–12 weeks</p> <p>Maternal age Mean 37 years (35–45)</p> <p>Drop-outs Failed quality control n=1 No karyotype results n=34</p>	<p>Platform NIPT Not specified</p> <p>Reference test Karyotype</p>	<p>Monosomy X TP 0 FP 1 FN 1 (mosaic 45X/47XXX) TN 176</p> <p>XXY TP 0 FP 0 FN 1 (mosaic 45X/47XXX) TN 177</p> <p>XXX TP 0 FP 0 FN 1 TN 177</p>	<p>Moderate</p> <p>Commercial partner Berry Genomics Co, Ltd</p>

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First author Year Reference Country	Study design	Population	Index test platform Reference test	Results for diagnosed trisomy or sex chromosome aneuploidies	Study quality Comments
Yao 2014 [27] China	<p>Study design Retrospective analysis, cohort study</p> <p>Not blinded</p> <p>Time of study June 2011 to December 2012</p>	<p>Population High risk for fetal aneuploidy n=5 950 (analysed by DNA sequencing) n=24 (karyotyping) n=5 466 (pregnancy outcome)</p> <p>Samples Not biobanked samples</p> <p>Inclusion criteria Singleton pregnancy Pre-test counseling</p> <p>Exclusion criteria Not specified</p> <p>Gestational age at sampling Mean 19 weeks</p> <p>Maternal age Mean 30 years</p> <p>Drop-outs Lost to follow-up, pregnancy outcome n=408 Of the 33 with a positive NIPT result for SCA 8 declined invasive confirmatory test One karyotype which was indicated as monosomy X failed</p>	<p>Platform NIPT Not specified</p> <p>Reference test Karyotyping or pregnancy outcome</p>	<p>Monosomy X (7 identified with NIPT, 5 analysed with karyotype) TP 2 FP 3 Lost to follow-up: 2</p> <p>XXX (14 identified with NIPT, 10 analysed with karyotype) TP 7 FP 3 Lost to follow-up: 4</p> <p>XXY (9 identified with NIPT, 7 analysed with karyotype) TP 3 FP 4 Lost to follow-up: 2</p> <p>XYY (3 identified with NIPT, 2 analysed with karyotype) TP 1 FP 1 Lost to follow-up: 1</p>	<p>Moderate</p> <p>Commercial partner BGI-Shenzhen</p>

AMA = Advanced maternal age; **CI** = Confidence interval; **CMA** = Chromosomal microarray analysis; **CNV** = Copy number variations; **DNA** = Deoxyribonucleic acid; **FISH** = Fluorescent in situ hybridization; **FN** = False negative; **FP** = False positive; **ITT** = Intention to treat; **MPSS** = Massive parallel shotgun sequencing; **n** = Number; **NIPT** = Non-invasive prenatal testing; **QF-PCR** = Quantitative fluorescence polymerase chain reaction; **SCA** = Sex chromosome; **SNP** = Single nucleotide polymorphism; **T-MPS** = Targeted massive parallel sequencing; **TN** = True negative; **TP** = True positive; **USS** = Ultrasound screening

Table 11.2 Studies with moderate risk of bias investigating Next-generation sequencing on microdeletion/duplications or whole genome coverage.

First author Year Reference Country	Study design	Population	Index test platform Reference test Verification test	Results	Study quality Comments
Chen 2013 [19] China	<p>Study design Cohort study</p> <p>Blinded</p> <p>Time of study Not reported</p>	<p>Population Total n=1 451 140 control samples with normal karyotype to develop algorithm Tested n=1 311</p> <p>Samples Biobanked NIPT</p> <p>Gestational age at sampling Mean 21 weeks (10–28)</p> <p>Inclusion criteria Women undergoing invasive prenatal diagnosis after maternal blood samples had been collected</p> <p>Exclusion criteria Not specified</p> <p>Maternal age Mean 32 years</p> <p>Drop-outs Not specified</p>	<p>Platform FCAPS</p> <p>Reference test Karyotype</p>	<p>Diagnosed CNV of >10 kb TP 3 FP 1</p> <p>Sensitivity: 100% Specificity: 99.92%</p>	<p>Moderate</p> <p>Commercial partner BGI-Shenzhen</p>

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Table 11.2 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test Verification test	Results	Study quality Comments
Helgeson 2015 [35] USA	<p>Study design Prospective cohort study</p> <p>Not Blinded</p> <p>Time of study October 2013 to October 2014</p>	<p>Population n=175 393</p> <p>n=123 096 analysed for deletions associated with 22q11.2 deletion syndrome, Cri du chat syndrome, Prader Willi and Angelman syndrome only</p> <p>Samples High risk for fetal aneuploidy (NIPT)</p> <p>Inclusion criteria High risk pregnancies including AMA, anomaly detected by USS, positive maternal serum screening, personal or family history</p> <p>Exclusion criteria Not specified</p> <p>Gestational age at sampling Not specified</p> <p>Maternal age Not specified</p> <p>Drop-outs Of the 55 that received a diagnosis based on the NIPT test, 2 were lost to follow-up. The number of dropouts and analysis failures from the entire cohort is not reported</p>	<p>Platform Not specified</p> <p>Verification CMA or FISH, clinical findings</p>	<p>Diagnoses n=55 1p36 deletion n=5 (TP 3, FP 1, 1 lost to follow-up)</p> <p>Wolf-Hirschhorn (4p16.3) n=1 (TP 1) Cri du chat (5p15) n=6 (TP 4, FP 2)</p> <p>Prader Willi/Angelman (15q11.2–q13) n=9 (TP 8, 1 positive based on clinical findings)</p> <p>22q11.2 deletion n=32 (both of maternal and fetal origin) (TP 23, 8 positive based on clinical findings 1 lost to follow-up)</p> <p>Jacobsen (11qter) n=1 (TP 1)</p> <p>Langer-Giedion (8q24.1) n=1 (1 positive based on clinical findings)</p>	<p>Moderate</p> <p>Commercial partner Authors employees at Sequenom laboratories</p>

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Table 11.2 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test Verification test	Results	Study quality Comments
Li 2016 [36] China	<p>Study design Retrospective cohort study</p> <p>Blinded</p> <p>Time of study September to December 2014</p>	<p>Population n=117 (total) n=117 (analysed)</p> <p>Samples NIPT on biobanked samples CMA on invasive samples CVS n=20 AF n=67 Cord blood n=38</p> <p>Inclusion criteria Women where invasive samples were analysed using CMA Anomaly detected by USS AMA Maternal anxiety</p> <p>Exclusion criteria Twin pregnancies Fetuses with common trisomies or SCA</p> <p>Gestational age at sampling Mean 21.3 weeks (12–34)</p> <p>Maternal age Mean 30 years (19–40)</p> <p>Drop-outs n=0</p>	<p>Platform Ion Proton sequencer (Life Technologies)</p> <p>Library Ion Plus Fragment Library Kit (Life Technologies)</p> <p>Reference CMA Cytoscan 750K Affymetrix reporting threshold 100kb</p>	<p>Diagnosed CNV ≥1 Mb TP 11 FP 4 TN 95 FN 7</p> <p>Sensitivity: 61.1% Specificity: 95%</p> <p>False positive rate 5%</p>	<p>Moderate</p> <p>Commercial partner None reported</p>

AMA = Advanced maternal age; AF = Amniotic fluid; CMA= Chromosomal microarray analysis; CNV = Copy number variations; CVS = Chorionic villus sampling; FCAPS = Fetal copy number analysis through maternal plasma sequencing; FISH = Fluorescent in situ hybridization; FN= False negative; FP = False positive; n = number; NIPT=Non-invasive prenatal testing; SCA = Sex chromosome aneuploidies; TN= True negative; TP = True positive; USS = Ultrasound screening

Table 11.3 Studies with high risk of bias investigating Next-generation sequencing on microdeletion/duplications or whole genome coverage.

First author Year Reference Country	Study design	Population	Index test platform Reference test Verification test	Results	Study quality Comments
Drury 2015 United kingdom [37]	Study design Proof of principle cohort study Blinded	Population n=24 (total) n=24 (analysed) Samples Invasive samples AF or CVS Inclusion criteria Anomaly detected by USS, including NT >3.5 Exclusion criteria Not sufficient DNA, abnormal karyotype, fetuses were CNVs had been identified by CMA Gestational age at sampling Not specified Maternal age Not specified Drop-outs Outcome of pregnancy: 2 cases lost to follow-up	Platform HiSeq 1 000 or HiSeq 2 500 (Illumina) or outsourcing to BGI Genomics Library TruSeq Exome (Illumina), Nextera Rapid exome kit (Illumina) or SureSelect All exon V (Agilent Biosystems) Reference Karyotype, CMA Verification Sanger sequencing	Diagnoses 5 definitive diagnoses (Milroy disease, hypophosphatasia, achondrogenesis type 2, Freeman-Sheldon syndrome and Baraitser- Winter Syndrome) 1 plausible diagnosis (orofacioidigital syndrome type 6)	Low Commercial partner Not reported

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Table 11.3 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test Verification test	Results	Study quality Comments
Fan 2012 [20] USA	Study design Proof of principle case control study	Population Case 1 Normal karyotype Case 2 22q11.2 microdeletion Maternal plasma and cord blood Gestational age at sampling Patient 1: First, second and third trimester Patient 2: Third trimester	Platform GAII and Hiseq (Illumina) Library SeqCap EZ Human Exome Kit v2.0 (Roche) Reference test Karyotype	Diagnoses 22q11.2 microdeletion	Low Commercial partner BGI-Shenzhen
Jensen 2012 [21] USA	Study design Proof of principle case control study Not Blinded Time of study Not reported	Population Cases 2 fetuses with known 22q11.2 microdeletions Control 14 fetuses low risk for chromosomal aberration Samples NIPT Gestational age at sampling Not specified Maternal age Not specified Drop-outs Not reported	Platform Hiseq 2 000 (Illumina) Library TruSeq (Illumina) Coverage 3.1-fold, 4.4-fold Repeated 15.87-fold, 16.77-fold Reference test Karyotype one of the two cases only	Diagnoses 22q11.2 microdeletion Both affected fetuses were detected	Low Commercial partner Sequenom laboratories

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Table 11.3 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test Verification test	Results	Study quality Comments
Wapner 2014 [23] USA	Study design Proof of principle case control study Blinded	Population n=469 (total) Cases n=6 Control n=352 Artificial DNA n=111 (both cases and controls) Samples Biobanked maternal plasma or artificial DNA Drop-outs Did not pass quality control n=23	Platform PCR based, using SNPs, NATUS algorithm	Diagnoses 1p36 microdeletion n=1 TP 1, FP 0 Cri du chat (5p15) n=24 TP 24, FP 1 Prader Willi/Angelman (15q11.2–q13) n=36 TP 36, FP 0 22q11.2 deletion n=46 TP 45, FP 3	Low Commercial partner Natera

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Table 11.3 continued

First author Year Reference Country	Study design	Population	Index test platform Reference test Verification test	Results	Study quality Comments
Yu 2013 [18] Hong Kong	<p>Study design Proof of principle case control study</p> <p>Not blinded</p>	<p>Samples 6 cases (3 post-invasive, 3 pre-invasive)</p> <p>Case 1 Post-invasive, cordocentesis 22q11.2 microdeletion, FISH</p> <p>Case 2 Post-invasive, cordocentesis 22q11.2 microdeletion, FISH</p> <p>Case 3 Post-invasive amniocentesis 22q11.2 microdeletion, QF-PCR and FISH</p> <p>Case 4 Pre-invasive Chorionic villus sampling 22q11.2 microduplication (2.4 Mb) Array CGH</p> <p>Case 5 Pre-invasive Amniocentesis 22q11.2 microduplication (2.4 Mb) Array CGH</p> <p>Case 6 Pre-invasive amniocentesis 3q29 microduplication (5.1 Mb); 4q32.1-q35.2 microdeletion (32.9 Mb) Array CGH</p> <p>8 controls</p>	<p>Platform HiSeq 2 000 (Illumina)</p> <p>Library TruSeq (Illumina)</p> <p>Coverage 3 Mb Resolution</p> <p>Reference test Karyotype CMA FISH QF-PCR</p>	<p>Diagnoses All 6 cases detected</p>	<p>Low</p> <p>Commercial partner None reported</p>

AF = Amniotic fluid; CGH = Comparative genomic hybridization; CMA = Chromosomal microarray analysis; CNV = Copy number variations; CVS = Chorionic villus sampling; DNA = Deoxyribonucleic acid; FISH = Fluorescent in situ hybridization; FP = False positive; Mb = Megabases; n = Number; NIPT = Non-invasive prenatal testing; NT = Nuchal translucency; PCR = Polymerase chain reaction; QF-PCR = Quantitative fluorescence-polymerase chain reaction; SNP = Single nucleotide polymorphism; TP = True positive; USS = Ultrasound screening

12 References

1. SBU. Fosterdiagnostik med mikroarray för utökad analys av kromosomer. Stockholm: Statens beredning för medicinsk och social utvärdering (SBU); 2016. SBU-rapport nr 246. ISBN 978-91-85413-89-8.
2. SBU. Utvärdering av metoder i hälso- och sjukvården: En handbok. Andra upplagan 2014. Stockholm: Statens beredning för medicinsk utvärdering (SBU). Hämtad från <http://www.sbu.se/metodbok>. 2015-12-30.
3. SBU. Analys av foster-DNA i kvinnans blod: icke-invasiv fosterdiagnostik (NIPT) för trisomi 13, 18 och 21. Stockholm: Statens beredning för medicinsk utvärdering (SBU); 2015. SBU Alert-rapport nr 2015-03. ISSN 1652-7151. <http://www.sbu.se>
4. SMER. Rapport 2015:1. Analys av foster-DNA i kvinnans blod: Icke invasiv fosterdiagnostik (NIPT) för trisomi 13, 18 och 21 – etiska aspekter. 2015.
5. Sunden B. Obstetric diagnosis with ultrasound. *Ultrasonics* 1967;5:67-71.
6. SFOG Sffog. Summarisk rapport för verksamhetsåret 2013. Hämtad från: https://www.sfog.se/media/193032/summarisk_rapport_sfog_2013.pdf. 2015-11-12. 2013.
7. SFOG Sffog. Årsrapport 2013. Graviditetsregistret. Hämtat från: <https://www.medscinet.com/GR/dokumentarkiv.aspx> 2015-11-12. 2013.
8. SBU. Analys av foster-DNA i kvinnans blod: icke-invasiv fosterdiagnostik för blodgrupps- eller könsbestämning. Stockholm: Statens beredning för medicinsk utvärdering (SBU); 2011. SBU Alert-rapport nr 2011-07. ISSN 1652-7151. <http://www.sbu.se>.
9. Orphanet. The portal for rare diseases and orphan drugs. Hämtat från <http://www.orpha.net/consor/cgi-bin/index.php>. 2015-12-03. 2015.
10. Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean L, et al. GeneReviews. Hämtad från <http://www.ncbi.nlm.nih.gov/books/NBK1116/>. 2015-12-03. 2015.
11. Socialstyrelsen. Kunskapsdatabas om ovanliga diagnoser. Hämtat från: <http://www.socialstyrelsen.se/ovanligadiagnoser>. 2015-11-19.
12. Socialstyrelsen. Fosterskador och kromosomavvikelser 2012. Hämtad från

- <http://www.socialstyrelsen.se/publikationer2013/2013-11-25>. 2013.
13. Petersen OB, Vogel I, Ekelund C, Hyett J, Tabor A. Potential diagnostic consequences of applying non-invasive prenatal testing: population-based study from a country with existing first-trimester screening. *Ultrasound Obstet Gynecol* 2014;43:265-71.
 14. Akolekar R, Beta J, Picciarelli G, Ogilvie C, D'Antonio F. Procedure-related risk of miscarriage following amniocentesis and chorionic villus sampling: a systematic review and meta-analysis. *Ultrasound Obstet Gynecol* 2015;45:16-26.
 15. Simpson JL. Invasive procedures for prenatal diagnosis: any future left? *Best Pract Res Clin Obstet Gynaecol* 2012;26:625-38.
 16. SFOG Sffog. Förslag till SFOG riktlinjer för fosterdiagnostik med NIPT, non invasive prenatal test. Preliminär version som presenterades under SFOG-veckan i Varberg 2015 av ULTRA-ARG. Hämtat från <https://www.sfog.se/start/rad-riktlinjer/sfog-riktlinjer/> 2015-11-24.
 17. Pergament E, Cuckle H, Zimmermann B, Banjevic M, Sigurjonsson S, Ryan A, et al. Single-nucleotide polymorphism-based non-invasive prenatal screening in a high-risk and low-risk cohort. *Obstet Gynecol* 2014;124:210-8.
 18. Yu SC, Jiang P, Choy KW, Chan KC, Won HS, Leung WC, et al. Noninvasive prenatal molecular karyotyping from maternal plasma. *PLoS One* 2013;8:e60968.
 19. Chen S, Lau TK, Zhang C, Xu C, Xu Z, Hu P, et al. A method for noninvasive detection of fetal large deletions/duplications by low coverage massively parallel sequencing. *Prenat Diagn* 2013;33:584-90.
 20. Fan HC, Gu W, Wang J, Blumenfeld YJ, El-Sayed YY, Quake SR. Non-invasive prenatal measurement of the fetal genome. *Nature*. 2012;487:320-4.
 21. Jensen TJ, Dzakula Z, Deciu C, van den Boom D, Ehrich M. Detection of microdeletion 22q11.2 in a fetus by next-generation sequencing of maternal plasma. *Clin Chem* 2012;58:1148-51.
 22. Bianchi DW, Platt LD, Goldberg JD, Abuhamad AZ, Sehnert AJ, Rava RP. Genome-wide fetal aneuploidy detection by maternal plasma DNA sequencing. *Obstet Gynecol* 2012;119:890-901.
 23. Wapner RJ, Babiarez JE, Levy B, et al. Expanding the scope of noninvasive prenatal testing: detection of fetal microdeletion syndromes. *Am J Obstet Gynecol* 2015; 212 (3) 332.e1-332.e9.
 24. Song Y, Huang S, Zhou X, Jiang Y, Qi Q, Bian X, et al. Non-invasive prenatal testing for fetal aneuploidies in the first trimester of pregnancy. *Ultrasound Obstet Gynecol* 2015;45:55-60.
 25. Song Y, Liu C, Qi H, Zhang Y, Bian X, Liu J. Noninvasive prenatal testing of fetal aneuploidies by massively parallel sequencing in a prospective Chinese population. *Prenat Diagn* 2013;33:700-6.
 26. Lau TK, Cheung SW, Lo PS, Pursley AN, Chan MK, Jiang F, et al. Non-invasive prenatal testing for fetal chromosomal abnormalities by low-coverage whole-genome sequencing of maternal plasma DNA: review of 1982 consecutive cases in a single center. *Ultrasound Obstet Gynecol* 2014;43:254-64.
 27. Yao H, Jiang F, Hu H, Gao Y, Zhu Z, Zhang H, et al. Detection of fetal sex chromosome aneuploidy by massively parallel sequencing of maternal plasma DNA: initial experience in a Chinese hospital. *Ultrasound Obstet Gynecol* 2014;44:17-24.
 28. Comas C, Echevarria M, Rodriguez MA, Prats P, Rodriguez I, Serra B. Initial experience with non-invasive prenatal testing of cell-free DNA for major chromosomal anomalies in a clinical setting. *J Matern Fetal Neonatal Med* 2015;28:1196-201.
 29. Porreco RP, Garite TJ, Maurel K, Marusiak B, Obstetrix Collaborative Research N, Ehrich M, et al. Noninvasive prenatal screening for fetal trisomies 21, 18, 13 and the common sex chromosome aneuploidies from maternal blood using

- massively parallel genomic sequencing of DNA. *Am J Obstet Gynecol.* 2014;211(4):365 e1-12.
30. Jiang F, Ren J, Chen F, Zhou Y, Xie J, Dan S, et al. Noninvasive Fetal Trisomy (NIFTY) test: an advanced noninvasive prenatal diagnosis methodology for fetal autosomal and sex chromosomal aneuploidies. *BMC Med Genomics* 2012;5:57.
 31. Liang D, Lv W, Wang H, Xu L, Liu J, Li H, et al. Non-invasive prenatal testing of fetal whole chromosome aneuploidy by massively parallel sequencing. *Prenat Diagn* 2013;33:409-15.
 32. Mazloom AR, Dzakula Z, Oeth P, Wang H, Jensen T, Tynan J, et al. Noninvasive prenatal detection of sex chromosomal aneuploidies by sequencing circulating cell-free DNA from maternal plasma. *Prenat Diagn* 2013;33:591-7.
 33. Nicolaidis KH, Syngelaki A, Gil M, Atanasova V, Markova D. Validation of targeted sequencing of single-nucleotide polymorphisms for non-invasive prenatal detection of aneuploidy of chromosomes 13, 18, 21, X, and Y. *Prenat Diagn* 2013;33:575-9.
 34. Shaw SW, Hsiao CH, Chen CY, Ren Y, Tian F, Tsai C, et al. Noninvasive prenatal testing for whole fetal chromosomal aneuploidies: a multicenter prospective cohort trial in Taiwan. *Fetal Diagn Ther* 2014;35:13-7.
 35. Hume JH, Wardrop J, Boomer T, et al. Clinical outcome of subchromosomal events detected by whole-genome non-invasive prenatal testing. *Prenat Diagn.* 2015;35(10):999–1004.
 36. Li R, Wan J, Zhang Y, Fu F, Ou Y, Jing X, et al. Detection of fetal copy number variations by noninvasive prenatal testing for common aneuploidies. *Ultrasound Obstet Gynecol.* 2016;47:53–7.
 37. Drury S, Williams H, Trump N, Boustred C, Lench N, Scott RH, et al. Exome sequencing for prenatal diagnosis of fetuses with sonographic abnormalities. *Prenat Diagn* 2015;35:1010-7.
 38. Agatisa PK, Mercer MB, Leek AC, Smith MB, Philipson E, Farrell RM. A first look at women's perspectives on noninvasive prenatal testing to detect sex chromosome aneuploidies and microdeletion syndromes. *Prenat Diagn* 2015;35:692-8.
 39. Bunnik EM, de Jong A, Nijssingh N, de Wert GM. The new genetics and informed consent: differentiating choice to preserve autonomy. *Bioethics* 2013;27:348-55.
 40. Dondorp WJ, de Wert GM. The 'thousand-dollar genome': an ethical exploration. *Eur J Hum Genet* 2013;21 Suppl 1:S6-26.
 41. Netzer C, Schmitz D, Henn W. To know or not to know the genomic sequence of a fetus. *Nat Rev Genet* 2012;13:676-7.
 42. Dondorp W, Sikkema-Raddatz B, de Die-Smulders C, de Wert G. Arrays in postnatal and prenatal diagnosis: An exploration of the ethics of consent. *Hum Mutat* 2012;33:916-22.
 43. Juth N. Genetic Information-Values and Rights. The morality of presymptomatic genetic testing. *Acta philosophica Gothoburgensia/Acta Universitatis Gothoburgensis*; 2005. ISBN 91-7346-534-8.
 44. Munthe C. The Moral Roots of Prenatal Diagnosis: Ethical Aspects of the Early Introduction and Presentation of Prenatal Diagnosis in Sweden, Hämtad från: <http://bit.ly/1CbsFnx> Studies in Research Ethics, Gothenburg. 1996.
 45. SMER. Rapport: Fosterdiagnostik – Etisk analys för diagnostik med foster-DNA, hämtad från: <http://www.smer.se/wp-content/uploads/2012/05/Rapport-Fosterdiagnostik-Etisk-analys-for-diagnostik-med-foster-DNA.pdf>. 2011.
 46. Lalatta F, Tint GS. Counseling parents before prenatal diagnosis: do we need

- to say more about the sex chromosome aneuploidies? *Am J Med Genet A* 2013;161a:2873-9.
47. Blackburn HL, Schroeder B, Turner C, Shriver CD, Ellsworth DL, Ellsworth RE. Management of incidental findings in the era of next-generation sequencing. *Curr Genomics* 2015;16:159-74.
 48. Green RC, Berg JS, Grody WW, Kalia SS, Korf BR, Martin CL, et al. ACMG recommendations for reporting of incidental findings in clinical exome and genome sequencing. *Genet Med* 2013;15:565-74.
 49. ACMG policy statement: updated recommendations regarding analysis and reporting of secondary findings in clinical genome-scale sequencing. *Genet Med* 2015;17:68-9.
 50. Riedijk S, Diderich KEM, van der Steen SL, Govaerts LCP, Joosten M, Knapen MFCM, et al. The psychological challenges of replacing conventional karyotyping with genomic SNP array analysis in prenatal testing. *J Clin Med* 2014;3:713-23.
 51. Simonato L, Niklas Juth, Christian Munthe: The ethics of screening in healthcare and medicine: serving society or serving the patient? *Theor Med and Bioeth* 2015;36:243-5.
 52. Gil MM, Revello R, Poon LC, Akolekar R, Nicolaides KH. Clinical implementation of routine screening for fetal trisomies in the UK NHS: cell-free DNA test contingent on results from first-trimester combined test. *Ultrasound Obstet Gynecol* 2016;47:45-52.